



Preliminary screening of active compounds with the evaluation of the anti-*Pseudomonas aeruginosa* bacteria activity of ashwagandha (*Withania Somnifera*) leaves extracted by different types of solvents

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ABSTRACT

Ashwagandha (*Withania Somnifera* L.) plant is a medicinal plant used to treat various ailments in Iraq by herbalists, the pharmacological evaluation of its leaves despite the rich pharmacological potential of its roots was not done, in this re- search, the phytochemicals from the Ashwagandha (*Withania Somnifera* L.) leaves were extracted by different types solvents (carbon tetrachloride, chloroform, ethanol and water respectively), presence flavonoids, tannins, saponin, steroids in water and ethanol extracts while the saponin and steroids only were present in chloroform and carbon tetrachloride extracts, the water and ethanolic extracts were found to be good inhibitor or good active against *Pseudomonas aeruginosa* bacteria, and this activity may be belong to polyphenolic compounds such as flavonoids and tannins compounds, and by comparing the GC/MS analysis chromatogram of extracted sample with NIST database entries, identification compounds by peak value of the unknown compound in chromatogram. In chloroform extract among the 16 phytochemicals identified, 1,2,3,5- Cyclohexanetetrol is the most abundant compound present followed by n-Hexadecanoic acid (Palmitic acid), then Acetic acid, 3,5-Dihydroxy-6-methyl-2,3-dihydro-4H-pyran-4-one, 5-Hydroxy methyl furfural, Caryophyllene oxide, α -Tocopherol- β -D-mannosidic respectively, other compounds revealed are n-Decanoic acid, Formic acid, cis-3-Hexenylpyruvate, 4-Octyl tetradecyl Phthalate ester, etc. In ethanol extract among the 21 phytochemicals identified, O- Butyl hydroxylamine is the most abundant compound present followed by Hexanoic acid ethyl ester, then Oleyl alcohol, trifluoroacetate, Oleic acid, etc.

Keywords: *Withania somnifera* L., Leaves, *Pseudomonas aeruginosa*, GC/MS, natural

INTRODUCTION

The herbal medicine had pass used in the past decades is to thrust heed to wild cultivated natural plants and to return to what is known as traditional medicine or herbal medicine (Arab medicine), which was used for previous centuries in Iraq mainly for medicine treatment and health care [1]. Ashwagandha (*Withania somnifera* L.) plant belongs to the Solanaceae family, it is a wild plant grown in semi-arid mild regions in the south and mid-Iraq [2].

The different parts of this plant contain different active compounds with a high pharmaceutical value [3,4]. Ashwagandha is an anoint medicinal herb with multiple health benefits. The roots crud and extract can reduce anxiety and stress, help fight depression, boost fertility and testosterone in men, and boost brain function [5]. In the Ayurvedic and Greek systems, the leaves of the plant are used for tumor and tubercular glands. Leaves are bitter in taste and used as an anthelmintic [1,2] [6,7], and anticholinesterase activities [8]. The root decoction boiled with milk and ghee is recommended for the treatment of sterility in women. while roots are only used in the treatment of constipation, senile debility, rheumatism, general debility, nervous exhaustion, loss of memory, excess spermatorrhoea in men, and loss of muscular energy [4]. Previous research has also shown that it is anti-stress, neuroprotective, anti-arthritic, anti-tumor, analgesic, and anti-inflammatory (maintaining health).[9], and indicate that *W. somnifera* possesses important bioactive properties which could be attributed to the high amounts of phenolic compounds.[10].

Pseudomonas aeruginosa is a type of germ that can cause infections in humans, mostly in hospital patients, it causes pneumonia [11]. Many *Pseudomonades aeruginosa* isolates are resistant to a large range of antibiotics and many demonstrate additional resistance to term unsuccessful treatment [12]. *Withania somnifera* L. has been widely studied for therapeutic purposes and possesses several pharmacological properties, including antioxidative, analgesic, anti-ulcerative, antiepileptic, and antibacterial properties [13,14].

The leaves of Ashwagandha alcoholic extract have the potential to induce senescence-like growth arrest and differentiation in glioma cells, these led to the formulation of a unique combination formula of alcoholic extract components that caused enhanced differentiation of glial cells [15], in other way of Ashwagandha leaf extract, cholinergic properties by provided biochemical and molecular evidence during brain disorders associated with cholinergic dysfunction [16], and quickly reduces Ag⁺ to Ag⁰ and enhances synthesis of silver nanoparticles with highly an-ti-microbial activity.[17], and trimethylene glycol is an active sleep-inducing component of Ashwagandha leaves and could potentially be useful for insomnia therapy[18], so, *W. somnifera* is a rich source of withanolides and other bioactive constituents, which can be used as a safe drug for various chronic diseases due to the minimal side effects in various researches pre-clinical. These researches are interesting and signify that more clinical trials should be conducted to prove the efficacy and other potential therapeutic effects in human settings [19,20,21].

The goal of this research included, *Withania somnifera* L. leaves extraction, and preliminary screening of the main active compound using different solvents such as carbon tetrachloride, chloroform, ethanol 80%, and water (deionized W.) respectively. Also, the aim included evaluation the of all extracts against *Pseudomonas aeruginosa* bacteria, the present

work aimed to identify the chemical compounds present in ethanol and chloroform extracts using GC- MS analyses, and this research contributes to the chemotaxonomic characterization of ethanol and chloroform extracts and validates these compounds as a potential source of natural antimicrobial and bacteria.

Materials and Methods

1.1. Plant material

The leaves of the plant *Withania somnifera* L. were collected from Al-Saidiya District, Baghdad, Iraq. The plant material was collected during April and washed with tap water to remove the debris, dried for two weeks at room temperature in the shade. The dried powder was packed in tightly closed containers and stored for phytochemical and antibacterial studies.

1.2. Leaves Extraction

Leaves powder of *Withania somnifera* L. 20 g was defatting with carbon tetrachloride (200mL) by Soxhlet apparatus till exhaustion (for 8hs) then the material was extracted by different solvents including chloroform, ethanol 80%, and water (deionized W.). The carbon tetrachloride, chloroform, and ethanolic extracts were dried by rotary evaporator while water (deionized W.) extract was dried by lyophilized apparatus.[12]

1.3. Chemical Tests

Detection of the main active compounds using chemical substances in general.

1.3.1. Flavonoids Test

Ethanolic potassium hydroxide (2mL) was added to (1mL) of extract of leaves. If flavonoid compounds are present in the extract a result yellow color is detected.

1.3.2. Terpenoids Test.

In a test tube mix (5mL) aqueous leaves extract with (2mL) chloroform, then add carefully (3mL) of concentrated sulphuric acid to the mixture to form a layer. If the terpenoid constituents are present, an interface with a reddish-brown color is found.

1.3.3. Saponin Test.

Powdered (1g) leaves sample was boiled together with (10mL) D.W. and then filtered, in a test tube mixed (5mL) of the filtered sample with (5mL) of D.W. and shaken vigorously to get a steadily permanent fourth. The formation of an emulsion by mixing the persistent forth with olive oil (3 drops) indicates the presence of saponins.

1.3.4. Tannins Test.

In a test tube (0.5g) of powdered plant, the sample is boiled with (20mL) of D.W. and then filtered, added to the filtered sample (0.1%) ferric chloride, if the presence of tannins that show observed brownish-green or a blue-black coloration.

1.3.5. Steroids Test.

In the test tube ethanolic leaves extract add (1mL) chloro- form, (2-3mL) of acetic anhydride, and drops of concentration H₂SO₄. If the presence of steroids the upper layer in the test tube turns into a red sulphuric acid layer showing yellow with green fluorescence.

After the detection of the main active compounds using chemical substances in general, all extracts were dried under laboratory temperature, and different solvents to investigate of residual of the main active compounds, the solvents were used CCl₄. CHCl₃, EtOH 80%, and deionized water respectively.

1.4. IR Infrared (IR)

FTIR spectrophotometer is a valuable tool for functional groups identified present in the plant extract. I.R. spectral studies were carried out in the solid state from leaves extracts for solvents (water, ethanol 80%, chloroform, and carbon tetrachloride), as pressed KBr pellets, and using Shimadzu transform (FT-IR) spectrophotometer in the range of 400-4000/cm.

1.5. Mass Spectroscopy

To tell us to identify and separate the individual plant extract components by GC-MS instrument; What are plant extract components composed of? How much of each of the components are present in plant extract of chloroform and ethanolic? The identification of a compound in GC-MS is not only by comparing its retention time to a standard with (GC) only but also by using its mass spectrum, making this a tremendously powerful analytical tool.

1.6. Antibacterial study

The *Pseudomonas aeruginosa* bacteria used in this study were obtained from Al-Yarmouk Hospital. The antibacterial culture was collected from a solid medium and inserted in the petri dishes (that contain the nutrient agar) by using incubation loops, then incubated these petri dishes at 37 °C for 24 hours. After incubation, the cultures were ready for use in the experiment. The leaf plant extract's antibacterial activity was tested by the agar-well-diffusion method. The gel puncture method was done by using the sterile corn borer to make wells of 6mm size in the agar of each plate. The bacteria with DMSO (Dimethyl sulfoxide) were considered as negative control while the amoxicillin was considered as position control, then (1mL) of bacteria suspension was added in the wells in each plate. The water, ethanol 80%, chloroform, and carbon tetrachloride leaves extracts were tested for antibacterial activity against *Pseudomonas aeruginosa*.

Results and Discussion

Preliminary phytochemical examination of ashwagandha leaves extracts based on the polarity of different solvents, namely, water, ethanol 80%, chloroform, and carbon tetrachloride, respectively, detected the existence of flavonoids, tannins, saponins, and

steroids in abundance in water and ethanol 80% extracts, while the saponins and steroids are few in chloroform and carbon tetrachloride extracts. The summary of results is obtained in Table 1. These results referred to the water and ethanol 80% sol- -vents had more polarity percentage compared with chloroform and carbon tetrachloride.

Table1. Qualitative profile of the phytochemical found in leaves extracts of Ashwagandha.

| No. | Solvents | Flavor | Tan- <u>nine</u> | Saponi | Steroids |
|-----|-------------------|--------|---------------------|--------|----------|
| | | | <u>Noid</u> | | |
| 1 | Water | +++ | +++ | +++ | ++ |
| 2 | EtOH 80% | +++ | +++ | +++ | ++ |
| 3 | CHCl ₃ | - | - | + | + |
| 4 | CCl ₄ | - | - | + | + |

3.2 Identification IR Method of function groups

Spectra were measured in terms of T%, and the peaks at specific $\hat{U} \text{ cm}^{-1}$ (Wavenumber) were assigned by functional group bonding as per the reference given in the instrument manual of Shimadzu FTIR. The explanation of IR spectrum values of plant leaves extract with CCl₄, CHCl₃, EtOH, and Water are shown in Figure1, Figure2, Figure3, and Figure4, respectively, and the explanation of IIR spectrum values of plant Leaves extract with CCl₄, CHCl₃, EtOH, and Water as in the Table2, Table3, Table4. Table5, respectively.

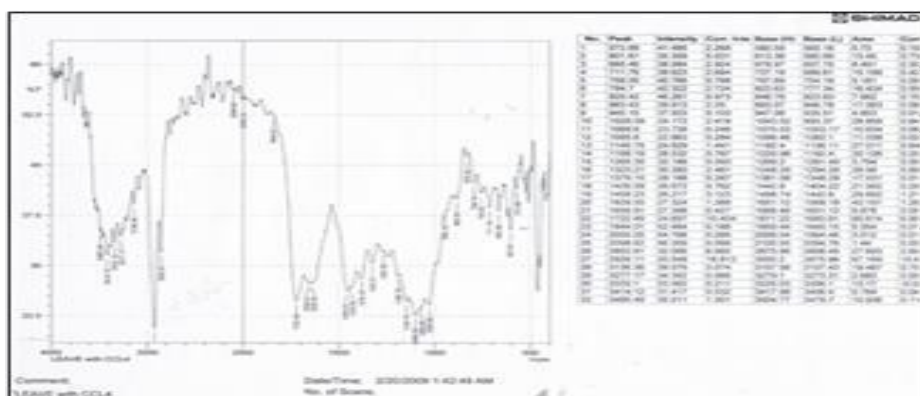


Figure 1. IR Spectrum of Leaves extract with CCl₄.

Table 2. Explanation of IR spectrum values of plant leaf extracts with CCl₄.

| No. | $\hat{U} \text{ cm}^{-1}$ | Transfer measure |
|-----|---------------------------|--------------------------|
| 1. | 3485.5, 3414 | N-H |
| 2. | 3333, 3277 | N-H amide |
| 3. | 3136, 2926 | O-H |
| 4. | 2852 | C-H Aldehyde hydrogen |

5. 2098 C≡C
6. 1722 C=O ester
7. 1656 C=O Amide
8. 1639, 1458 N-H Amide or Primary or Secondary amine
9. 1435, 1379, 1323 C-O-H
10. 1265, 1188, 1145 C-O ether

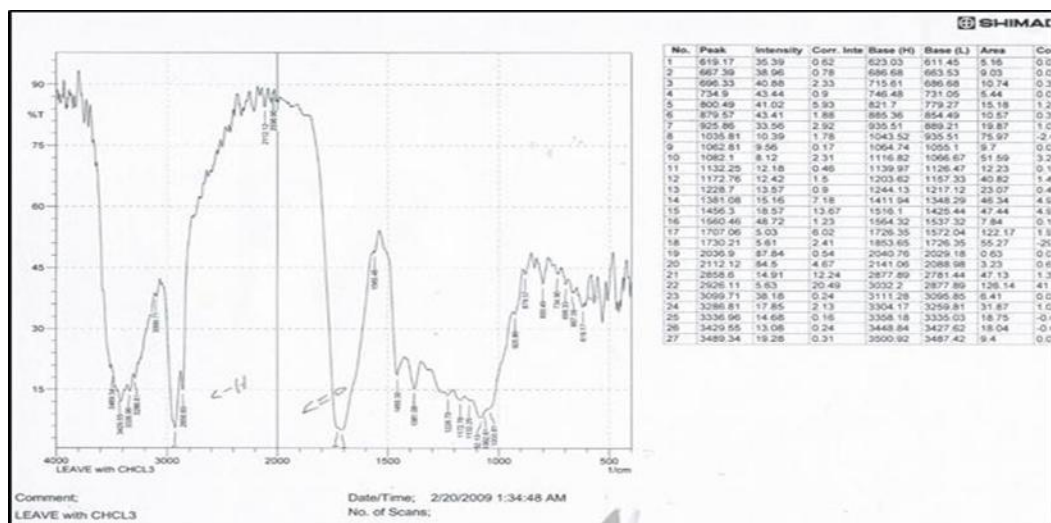


Figure 2. IR Spectrum of Leaves extract with CHCl₃.

Table 3. Explanation of IR spectrum values of plant leaves extract with CHCl₃.

| No. | \hat{U} cm ⁻¹ | Transfer measure |
|-----|----------------------------|----------------------------|
| 1. | 3489, 3429 | N-H |
| 2. | 3336, 3286 | N-H amide |
| 3. | 3099, 2926 | O-H |
| 4. | 2852 | C-H Aldehyde hydrogen |
| 5. | 2098 | C≡C |
| 6. | 1722 | C=O ester |
| 7. | 1656 | C=O Amide |
| 8. | 1639 | N-H Amide or Primary amine |
| 9. | 1458 | N-H Secondary amine |
| 10. | 1435, 1379, 1323 | C-O-H |
| 11. | 1265, 1188, 1145 | C-O ether |

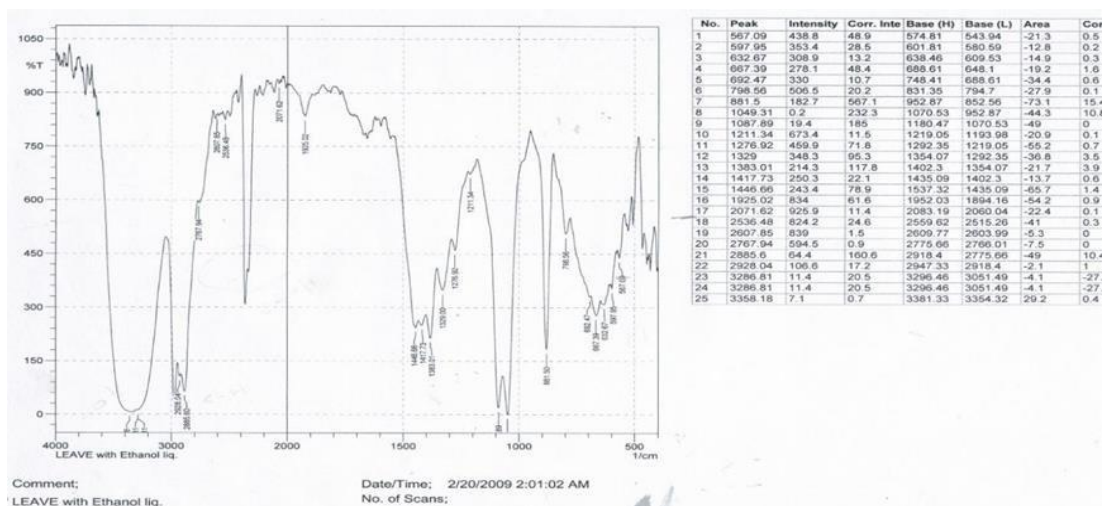
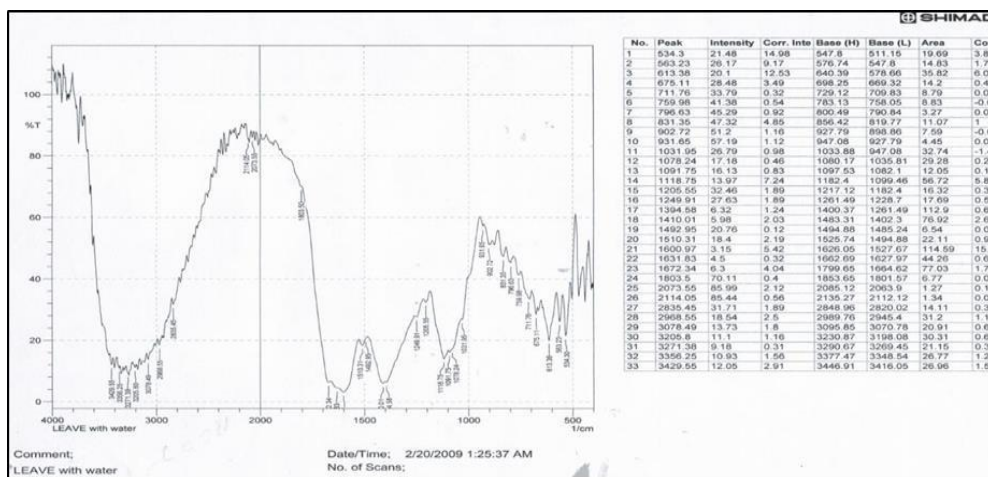


Figure 3. IR Spectrum of Leaves extract with EtOH.

Table 4. Explanation of IR spectrum values of plant leaves extract with EtOH Units.



| No. | \hat{U} cm-1 | Transfer measure |
|-----|------------------------------|---------------------|
| 1. | 3358 | \equiv C -H |
| 2. | 3286 | N-H amide |
| 3. | 2928, 2885, 2727, 2607, 2536 | O-H Carboxylic acid |
| 4. | 2071 | C \equiv C |
| 5. | 1417, 1383, 1329, 1276 | C-O-H |
| 6. | 1211, 1087, 1049 | C-O alcohol |
| 7. | 881, 796, 692, 667 | =C-H |

Figure 4. IR Spectrum of Leaves extract with H2O.

Table 5. Explanation of IR spectrum values of plant leaves extract with H₂O.

| No. | $\tilde{\nu}$ cm-1 | Transfer measure |
|-----|------------------------------|-----------------------|
| 1. | 3429 | \equiv C-H |
| 2. | 3356 | N-H |
| 3. | 3271 | N-H amide |
| 4. | 3205, 3078, 2968 | O-H Carboxylic acid |
| 5. | 2835 | C-H Aldehyde hydrogen |
| 6. | 2114, 2073 | C \equiv C |
| 7. | 1672, 1631, | C=O Amide |
| 8. | 1600, 1510, 1492 | C=C |
| 9. | 1410, 1394, 1249 | C-O-H |
| 10. | 1205, 1118, 1091, 1078, 1031 | C-O Alcohol |
| 11. | 931, 902, 831, 796 | =C-H |

1.1. GC-Mass Spectroscopy Assay

The GC- Mass Spectroscopy Assay of Ashwagandha leaves from chloroform and ethanol extracts; In the chloroform extract, sixteen compounds were found through the results of GC- Mass Spectroscopy chromatogram analysis of the extract leaves, as shown in Figure 5 and Table 6, Learn about the compounds formula (MF), identification, and percentage in extract. Compound 1 as formic acid with MF [CH₂O₂] (m/z 46), constitutes 1.35% of chloroform extract; Compound 2 as Acetic acid with MF [C₂H₄O₂] (m/z 60), it constitutes 5.15% of chloroform extract; Compound 3 as 1-hydroxy 2-propanone with MF [C₃H₆O₂] (m/z 74), it constitutes 0.64% of chloroform extract; Compound 4 as ethenyl 2-Propenoate ester with MF [C₃H₆O₂] (m/z 98), it constitutes 0.26% of chloroform extract; Compound 5 as N, N, O-Tri- acetyl hydroxylamine with MF [C₆H₉NO₄] (m/z 159). it constitutes 0.35% of chloroform extract; Compound 6 as 3,5-dihydroxy-6-methyl- 2,3- dihydro - 4H-pyran-4-one with MF [C₆H₈O₄] (m/z 144), it constitutes 4.79% of chloroform extract; Compound 7 as 5-Hydroxy methyl furfural with MF [C₆H₆O₃] (m/z 126), it constitutes 3.68% of chloroform extract; Compound 8 as 1,1,3-trimethyl Cyclopentane with MF [C₈H₁₆] (m/z 112), it constitutes 0.35% of chloroform extract; Compound 9 as Caryophyllene oxide with MF [C₁₅H₂₄O] (m/z 220), it constitutes 2.43% of chloroform extract; Compound 10 as 1,2,3,5-Cyclohexanetetrol with MF [C₁₅H₂₄O] (m/z 148), it constitutes 64.16% of chloroform extract; Compound 11 as n-Decanoic acid with MF [C₁₀H₂₀O₂] (m/z 172), it constitutes 1.67% of chloroform extract; Compound 12 as n-Hexadecanoic acid with MF [C₁₆H₃₂O₂] (m/z 256), it constitutes 10.15% of chloroform extract; Compound 13 as cis-3-Hexenylpyruvate with MF [C₉H₁₄O₃] (m/z 170), it constitutes 1.30% of chloroform extract; Compound 14 as 1,2-Pentanediol with MF [C₃₀H₅₀O₄] (m/z 104). it constitutes 0.66% of chloroform extract; Compound 15 is 4-octyl tetradecyl Phthalate ester with MF [C₃₀H₅₀O₄] (m/z 474), constitutes 1.02% of chloroform extract; Compound 16 as α - Tocopherol- β -D-mannoside with MF [C₃₅H₆₀O₇] (m/z 592), it constitutes 1.02% of chloroform extract

Table 6. Compounds in Chloroform extract of Ashwagandha leaves from GC-MS

| - | Compound Name | Formula | M. mass | Area % | R.Time |
|-----|---|--|---------|--------|--------|
| 1. | Formic acid | CH ₂ O ₂ | 46 | 1.35 | 2.098 |
| 2. | Acetic acid | C ₂ H ₄ O ₂ | 60 | 5.15 | 2.226 |
| 3. | 1-Hydroxy 2-propanone | C ₃ H ₆ O ₂ | 74 | 0.64 | 2.434 |
| 4. | Ethenyl 2-Propenoate ester | C ₅ H ₆ O ₂ | 98 | 0.26 | 2.965 |
| 5. | N, N, O-Triacetyl hydroxylamine | C ₆ H ₉ NO ₄ | 159 | 0.35 | 3.242 |
| 6. | 3,5-Dihydroxy-6-methyl-2,3-dihydro-4H-pyran-4-one | C ₆ H ₈ O ₄ | 144 | 4.79 | 8.901 |
| 7. | 5-Hydroxy methyl furfural | C ₆ H ₆ O ₃ | 126 | 3.68 | 10.294 |
| 8. | 1,1,3-Trimethyl Cyclopentane | C ₈ H ₁₆ | 112 | 0.70 | 15.050 |
| 9. | Caryophyllene oxide | C ₁₅ H ₂₄ O | 220 | 2.43 | 15.245 |
| 10. | 1,2,3,5-Cyclohexanetetrol | C ₁₅ H ₂₄ O | 148 | 64.16 | 16.143 |
| 11. | n-Decanoic acid | C ₁₀ H ₂₀ O ₂ | 172 | 1.67 | 17.191 |
| 12. | n-Hexadecanoic acid | C ₁₆ H ₃₂ O ₂ | 256 | 10.15 | 19.370 |
| 13. | cis-3-Hexenylpyruvate | C ₉ H ₁₄ O ₃ | 170 | 1.30 | 21.110 |
| 14. | 1,2-Pentanediol | C ₃₀ H ₅₀ O ₄ | 104 | 0.66 | 21.303 |
| 15. | 4-Octyl tetradecyl Phthalate ester | C ₃₀ H ₅₀ O ₄ | 474 | 1.02 | 24.690 |
| 16. | α-Tocopherol-β-D-mannoside | C ₃₅ H ₆₀ O ₇ | 592 | 2.04 | 24.823 |

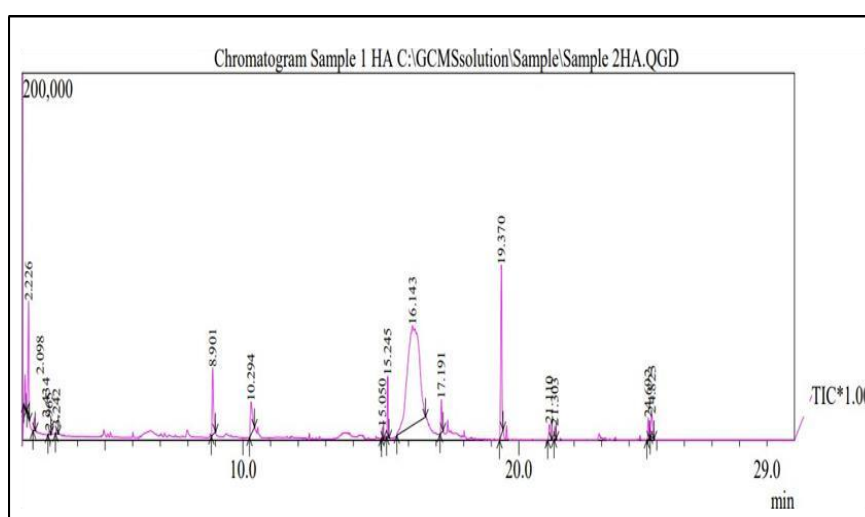


Figure 5. GC-MS Spectrum of Leaves extract with Chloroform.

In the ethanol extract of Ashwagandha leaves, twenty one compounds were found through the results of GC-Mass Spectroscopy chromatogram analysis, as show in Figure 6, and fourteen compounds are the largest percentage in the alcoholic leaves extract as show in Table 7, Compound 1 as Hexanoic acid, ethyl ester with MF [C₈H₁₆O₂] (m/z 144.2114), it constitutes 20.03% of ethanol extract; Compound 2 as O-Butyl hydroxylamine with MF [C₄H₁₁NO] (m/z 89.14), it constitutes 52.70% of ethanol extract; Compound 3 as N- ethyl propanamide with MF [C₅H₁₁NO] (m/z 101.15), it constitutes 0.17% of ethanol extract; Compound 4 as 1,3,6 Trioxocane with MF [C₅H₁₀O₃] (m/z 118.13), it constitutes 0.64% of ethanol extract; Compound 5 as Butanoic acid, ethyl ester with MF [C₆H₁₂O₂] (m/z 116.1583), it constitutes 0.20% of ethanol extract; Compound 6 as Oleyl alcohol, trifluoroacetate with MF [C₂₀H₃₅F₃O₂] (m/z 364.4859), it constitutes 7.36% of ethanol extract; Compound 7 as Cis- Vaccenic acid with MF [C₁₈H₃₄O₂] (m/z 282.4614), it constitutes 1.71% of ethanol extract; Compound 8 as Oleic acid with MF [C₁₈H₃₄O₂] (m/z 282.4614), it constitutes 5.65% of ethanol extract; Compound 9 as Cis 9-hexadecenal with MF [C₁₆H₃₀O] (m/z 238.4088), it constitutes 0.87% of ethanol extract; Compound 10 as 13-octadecenal, Z- with MF [C₁₈H₃₄O] (m/z 266.5), it constitutes 1.12% of ethanol extract; Compound 11 as (S)(+)-Z-13-Methyl 11-pentadecena- 1-ol acetate with MF [C₁₈H₃₄O₂] (m/z 282.4614), it constitutes 0.58% of ethanol extract; Compound 12 as (S)(+)-Z-13-Methyl 11-pentadecena-1-ol acetate with MF [C₁₈H₃₄O₂] (m/z 282.4614). it constitutes 1.49% of ethanol extract; Compound 13 as 7-Tetradecenal, Z- with MF [C₁₄H₂₆O] (m/z 210.3556). it constitutes 0.84% of ethanol extract; Compound 14 as Oxirane, tetradecyl- with MF [C₁₆H₃₂O] (m/z 240.4247), it constitutes 0.47% of ethanol extract; Compound 15 as Cis-13-Octadecenoic acid with MF [C₁₈H₃₄O₂] (m/z 282.4614), it constitutes 0.84% of ethanol extract; Compound 16 as 9-methyl-z, z-10,12-hexadecadien-1-ol acetate with MF [C₁₉H₃₄O₂] (m/z 294.4879), it constitutes 0.57% of ethanol extract; Compound 17 as 13-Octadecenal, Z- with MF [C₁₈H₃₄O] (m/z 266.5), it constitutes 1.02% of ethanol extract; Compound 18 as 9- Octadecenoic acid, Z-, methyl ester with MF [C₁₉H₃₆O₂] (m/z 296.4879), it constitutes 0.43% of ethanol extract; Compound 19 as Erucic acid with MF [C₂₂H₄₂O₂] (m/z 338.5677), it constitutes 0.10% of ethanol extract; Compound 20 as Oleic acid with MF [C₁₈H₃₄O₂] (m/z 282.4614), it constitutes 1.84% of ethanol extract; Compound 21 as Cis 9-hexadecenal with MF [C₁₆H₃₀O] (m/z 238.4088), it constitutes 1.36% of ethanol extract.

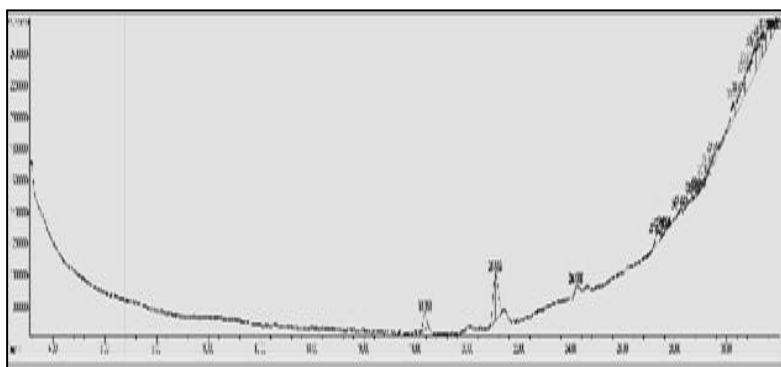


Figure 6. GC-MS Spectrum of Leaves Extract with Ethanol.

Table 7. Compounds in Ethanol extract of Ashwagandha leaves from GC-MS.

| No. | Compound Name | Formula | M. mass | Area % | RTime |
|-----|---|---|----------|--------|--------|
| 1. | Hexanoic acid, ethyl ester | C ₈ H ₁₆ O ₂ | 144.2114 | 20.03 | 18.351 |
| 2. | O-Butyl hydroxylamine | C ₄ H ₁₁ NO | 89.14 | 52.70 | 21.084 |
| 3. | N- ethyl- Propanamide | C ₅ H ₁₁ NO | 101.15 | 0.17 | 21.346 |
| 4. | 1,3,6 Trioxocane | C ₅ H ₁₀ O ₃ | 118.13 | 0.64 | 21.411 |
| 5. | Butanoic acid, ethyl ester | C ₆ H ₁₂ O ₂ | 116.1583 | 0.20 | 21.433 |
| 6. | Oleyl alcohol, trifluoroacetate | C ₂₀ H ₃₅ F ₃ O ₂ | 364.4859 | 7.36 | 27.317 |
| 7. | Cis-Vaccenic acid | C ₁₈ H ₃₄ O ₂ | 282.4614 | 1.71 | 27.357 |
| 8. | Oleic acid | C ₁₈ H ₃₄ O ₂ | 282.4614 | 5.65 | 27.401 |
| 9. | Cis 9-hexadecenal | C ₁₆ H ₃₀ O | 238.4088 | 0.87 | 29.109 |
| 10. | 13-octadecenal, Z- | C ₁₈ H ₃₄ O | 266.5 | 1.12 | 29.147 |
| 11. | (S)(+)Z-13-Methyl 11-pentadecena-1-ol acetate | C ₁₈ H ₃₄ O ₂ | 282.4614 | 0.58 | 29.190 |
| 12. | (S)(+)Z-13-Methyl 11-pentadecena-1-ol acetate | C ₁₈ H ₃₄ O ₂ | 282.4614 | 1.49 | 29.245 |
| 13. | 7-Tetradecenal, Z- | C ₁₄ H ₂₆ O | 210.3556 | 0.84 | 29.289 |
| 14. | Oxirane, tetradecyl- | C ₁₆ H ₃₂ O | 240.4247 | 0.47 | 29.370 |
| 15. | Cis-13-Octadecenoic acid | C ₁₈ H ₃₄ O ₂ | 282.4614 | 0.84 | 29.507 |
| 16. | 9-methyl-z,z-10,12-hexadecadien-1-ol acetate | C ₁₉ H ₃₄ O ₂ | 294.4879 | 0.57 | 29.567 |
| 17. | 13-Otadecenal, Z- | C ₁₈ H ₃₄ O | 266.5 | 1.02 | 29.692 |
| 18. | 9- Octadecenoic acid, Z-, methyl ester | C ₁₉ H ₃₆ O ₂ | 296.4879 | 0.43 | 29.741 |
| 19. | Erucic acid | C ₂₂ H ₄₂ O ₂ | 338.5677 | 0.10 | 29.769 |
| 20. | Oleic acid | C ₁₈ H ₃₄ O ₂ | 282.4614 | 1.84 | 29.878 |
| 21. | Cis 9-hexadecenal | C ₁₆ H ₃₀ O | 238.41 | 1.36 | 30.702 |

1.2. Antibacterial assay

The water and ethanolic extracts at 500 concerts showed more activity as anti-P. aeruginosa bacteria compared with chloroform and carbon tetrachloride, as shown in Figure 7. The inhibitor zones were 22,20 mm of water and ethanolic extracts while 15 and 12 mm of chloroform and carbon tetrachloride respectively. These results referred to more phenolic compounds in water and ethanol extracts such as flavonoids, tannin, and steroids which are considered antiseptic compounds. The inhibitory zones of water, ethanol, chloroform, and carbon tetrachloride extracts are presented in Table 8.

Table 8. The inhibition zone of Ashwagandha leaves extracts against *Pseudomonas aeruginosa* bacteria.

| Extract with | Water | Ethanol 80% | Chloroform | CCl4 | DMSO | Amoxicillin |
|----------------------|-------|-------------|------------|------|------|-------------|
| Inhibition Zone (mm) | 22 | 20 | 13 | 12 | Zero | 18 |

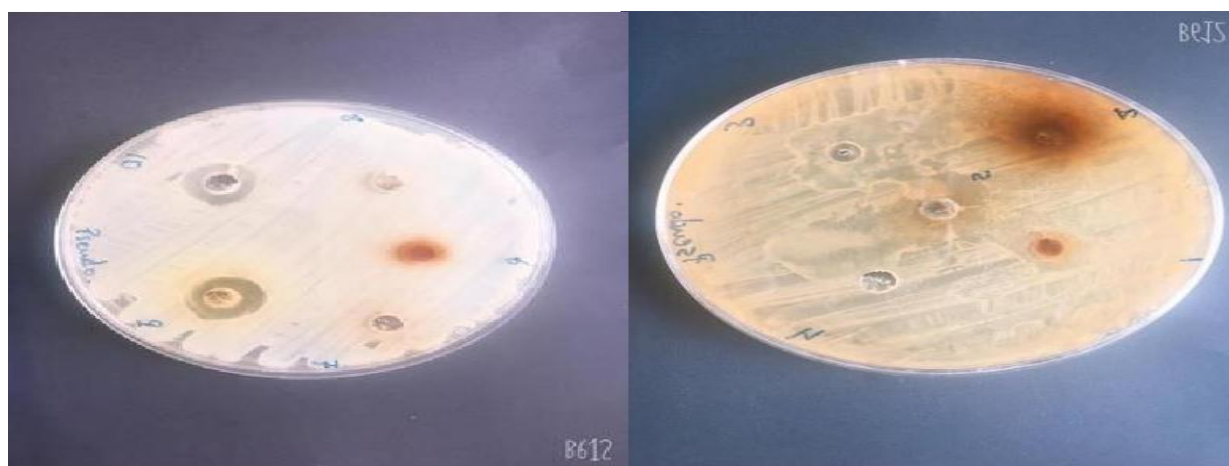


Figure 7. Showed anti-*P. aeruginosa* activity. 2- Leaves Extract with CCl₄, 5- Leaves Extract with water, 9- Leaves Extract with Ethanol, 10- Leaves Extract with Chloroform.

Flavonoids are synthesized by plants as a defense reaction against plant microbial infection, thus extracts from plants rich in flavonoids are used as bactericidal aid. They act by deactivating bacteria toxins or inhibiting the synthesis of bacterial DNA or they act by altering the outer bacterial cell membrane. The water and ethanolic extracts were found to be good inhibitors or good active against *Pseudomonas aeruginosa* bacteria, and this activity may belong to polyphenolic compounds such as flavonoids and tannins' compounds. As part of scientific research to extract bioactive natural products with novel structures from plants, we measured a phytochemical examination of the leaves of *W. somnifera* extract using- Mass Spectroscopy Assay, by comparing the chromatogram of GC-Mass Spectroscopy Assay peak value of the unknown compound with entries in NIST database, the identification of compounds was done. Among the 16 phytochemicals identified in chloroform extract, 1,2,3,5-Cyclohexanetetrol is the most abundant compound present (64.16%) in the extract, followed by n- Hexadecanoic acid (10.15%), then Acetic acid (5.15%), 3,5-Dihydroxy-6-methyl-2,3-dihydro-4H-pyran-4-one (4.79%), 5-Hydroxy methyl furfural (3.68%), Caryophyllene oxide (2.43%), α -Tocopherol- β -D-mannoside (2.04%), n-Decanoic acid (1.67%), Formic acid (1.35%), cis-3-Hexenylpyruvate (1.30%), 4-Octyl tetradecyl Phthalate ester (1.02%), 1,2-Pentanediol (0.66%), 1- Hydroxy 2-propanone (0.64%), N, N, O-Triacetyl hydroxylamine (0.35%), 1,1,3-Trimethyl Cyclopentane (0.35%), Ethenyl 2-Propenoate ester (0.26%). And

the 21 phytochemicals identified in ethanol extract, O-Butyl hydroxylamine is the most abundant compound present (52.70%) from ethanol extract, followed by Hexanoic acid, ethyl ester (20.03%), then Oleyl alcohol, trifluoroacetate (7.36%), Oleic acid, Cis 9-hexadecenal .est. An analysis of the leaves of this medicinal plant shows some compounds with rich pharmaceutical potential, which requires more scientific research experiments to identify and discover some new drugs that may be found in the leaves of this plant.

Conclusions

Four kinds of solvents were used to extract *Withania Somifera* L. leaves, the phytochemical investigation of leaves extract revealed the presence of important bioactive compounds such as flavonoids, tannins, saponin, and steroids.

The present study indicated that the plant leaves contain an antibacterial compound that can be further developed as phytomedicine for the therapy of infection.

2. Acknowledgments

The authors would like to thank the College of Pharmacy, Mustansiriyah University, Baghdad – Iraq for its support in the present work.

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